

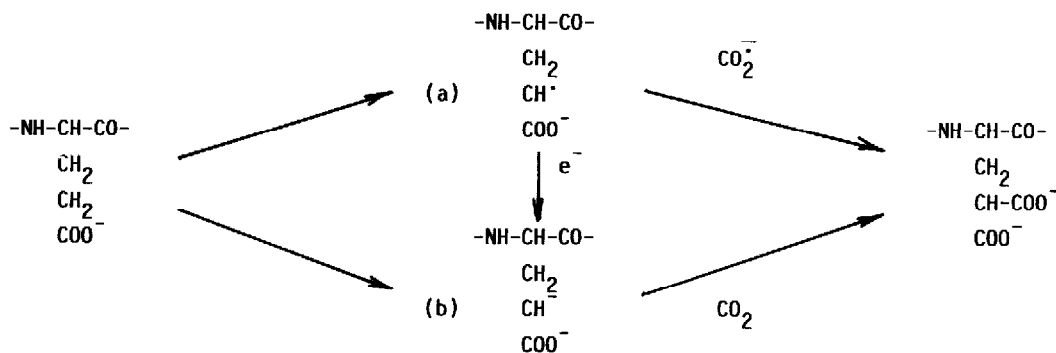
### HOMOLYTIC DECARBOXYLATION OF GLUTAMATE ANALOGUES

Anne Vidal-Cros, Sonia Bory, Michel Gaudry and Andrée Marquet

Laboratoire de Chimie Organique Biologique, associé au CNRS  
 Université Pierre et Marie Curie - 4, place Jussieu  
 75252 PARIS CEDEX 05, France

**Abstract :** The homolytic decarboxylation of a 3-cyclopropylglutamate derivative yields some rearrangement product whereas that of a 3-fluoroglutamate derivative yields the decarboxylation product without elimination. These results are discussed in relationship with the study of vitamin K-dependent carboxylation.

The mechanism of the post-ribosomal carboxylation of glutamyl residues catalyzed by vitamin K<sup>1,2</sup> is still obscure. Two intermediates can be *a priori* involved : a radical (a) and a carbanion (b).

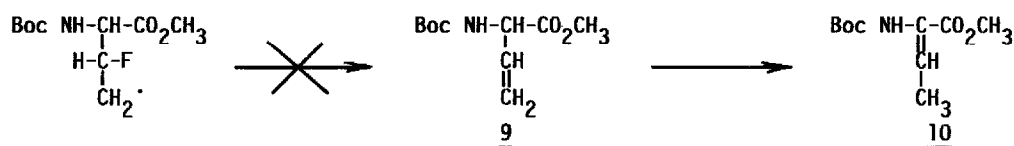


We synthesized two glutamate analogues designed to probe the occurrence of intermediate (a) or (b) during the enzymatic reaction. Here, we test the basis of our working hypotheses on the reactivity induced by the functionality of these analogues.

A positive argument for a radical would be the rearrangement of a cyclopropyl analogue<sup>3,4</sup>, for instance compound 1<sup>5</sup> during the enzymatic carboxylation. Here, we show that radical 2, generated from 1 by homolytic decarboxylation according to Barton<sup>8</sup>, actually rearranges : it reacts<sup>9</sup> with H<sup>•</sup> to yield the simple decarboxylation product 3<sup>10</sup> (49 %) and rearranges to give 4<sup>11</sup> (23 %).



In addition to decarboxylation products 7e and 7t<sup>19</sup>, we observed the formation of 8<sup>20</sup>. The low yield of decarboxylated material could be explained by H<sup>•</sup> capture competitively with the decarboxylation. This suggests that the presence of fluorine slows down the formation of a radical on the neighbouring carbon. Nevertheless, the main result for our discussion is that the dehydroproduct 10<sup>21</sup>, which could have been formed by F<sup>•</sup> elimination and isomerisation in the basic medium, was not observed.



The radicals possibly formed from 1 and 5 by the enzyme have an extra carboxyl group in  $\alpha$ . We should have studied the decarboxylation of the corresponding malonic acids. To our knowledge, Barton's method<sup>8</sup> has not been applied to malonic acids and our attempts to decarboxylate benzylmalonic acid failed. However there is, so far, no evidence, that an  $\alpha$  carboxyl group which has a stabilizing effect<sup>22</sup>, should favor the F<sup>•</sup> elimination.

We have studied recently the interaction of the carboxylase with 5 and observed that 5e yields the elimination product<sup>23</sup>. The results obtained here strongly suggest that a carbanion is involved. Work with the cyclopropyl derivative 1 is in progress.

ACKNOWLEDGEMENT : Y. HERVE is gratefully thanked for helpful advice concerning the decarboxylation procedure.

#### REFERENCES AND NOTES

1. J.W. SUTTIE, Ann. Rev. Biochem. (1985), 54, 450.
2. See in Current Advances in Vitamin K Research, J.W. SUTTIE (Ed). Elsevier (1988).
3. A.L.J. BECKWITH, G. MOAD, J. Chem. Soc., Perkin Trans 2 (1980), 1473.
4. a) J.E. BALDWIN, R.M. ADLINGTON, B.P. DOMAYNE-HAYMAN, G. KNIGHT, H.H. TING. J. Chem. Soc., Chem. Commun. (1987), 1661 ; A.J. CASTELLINO, T.C. BRUICE, J. Am. Chem. Soc. (1988), 110, 1313 and references cited therein.
5. Treatment of 2-t-butyloxycarbonylamino-3-methyleneglutarate 1-methyl-5-benzyl-diester<sup>6</sup> with diazomethane in excess in diethylether, in the presence of palladium acetate according to Suda<sup>7</sup> yielded 1-benyloxycarbonylmethyl-1-[t-butyloxycarbonylamino-methoxycarbonyl-methyl]-cyclopropane in 37% yield after column purification. Hydrogenolysis (H<sub>2</sub>, 10 % Pd/C, methanol) afforded 1. F = 135-136°C. <sup>1</sup>H NMR (CDCl<sub>3</sub>) 0.71 (bd, 4H, CH<sub>2</sub>-CH<sub>2</sub>), 1.42 (s, 9H, tBu), 2.42 (AB, 2H,  $\delta_A = 2.25$ ,  $\delta_B = 2.59$ , J<sub>AB</sub> = 16, CH<sub>2</sub>), 3.73 (s, 3H, OCH<sub>3</sub>), 3.85 (bm, 1H, CH), 5.10 (b signal, 1H, CO<sub>2</sub>H), 5.87 (b signal, 1H, NH).
6. Y.H. PAIK, P. DOWD, J. Org. Chem. (1986), 51, 2910.
7. M. SUDA, Synthesis (1981), 714.

8. D.H.R. BARTON, Y. HERVE, P. POTIER, J. THIERRY, J. Chem. Soc. Chem. Commun. (1984), 1298.
9. Decarboxylation general procedure : The acid (0.19 M in dry DMF, -15°C) was treated with N-methylmorpholine (1 eq), isobutylchloroformate (1 eq) and after 5 min with N-hydroxypyridine-2-thione (1,17 eq). After 20 min, 2-methyl-2-propanethiol (10 eq) was added and the mixture irradiated with two 100 W tungsten lamps for 1 h at 0°C. Decarboxylation of Boc Glu-OBz yields benzyl 2-t-butyloxycarbonylamino-butanoate (90 %).
10. 3.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ) 0.35-0.80 (m, H, cyclopropyl), 1.05 (s, 3H,  $\text{CH}_3$ ), 1.50 (s, 9H, tBu), 3.75 (s, 4H,  $\text{OCH}_3$  + H), 5.20 (m, 1H, NH).
11. 4.  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ) 0.90 (t, 3H,  $\text{CH}_3$ ,  $J = 7$ ), 1.50 (s, 9H, tBu), 2.15 (q, 2H,  $\text{CH}_2$ ,  $J = 7$ ), 3.80 (s, 3H,  $\text{OCH}_3$ ), 4.78 (d, 1H, CH,  $J = 8$ ), 5.08 (d, 2H,  $\text{CH}_2$ =), 5.30 (m, 1H, NH).
12. a) R. RANDO, Pharmacol. Rev. (1984), 36, 111 ; b) C. WALSH, Tetrahedron (1982), 38, 871.
13. J. STUBBE, S. FISH, R.H. ABELES, J. Biol. Chem. (1980), 255, 236.
14. A. VIDAL-CROS, M. GAUDRY, A. MARQUET, Biochem. J. (1985), 229, 675.
15. S. PATAI, "The Chemistry of the Carbon-Halogen Bond" (1973), Part 1, I. J. Wiley and Sons.
16. D.H.R. BARTON, N.K. BASU, R.H. HESSE, F.S. MOREHOUSE, M.M. PECHEI, J. Am. Chem. Soc. (1966), 88, 3016.
17. P. HERDEWIJN, R. PAUWELS, M. BABA, J. BALZARINI, E. DE CLERQ, J. Med. Chem. (1987), 30, 2131.
18. A. VIDAL-CROS, M. GAUDRY, A. MARQUET a) J. Org. Chem. (1985), 50, 3163 ; b) J. Org. Chem., (1989), 54, 498.
19. The reaction was performed under a  $\text{N}_2$  stream and the resulting  $\text{CO}_2$  bubbled in baryum hydroxide. The baryum carbonate was filtered off and weighted.
- 7 :  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ) *erythro* 1.42 (s, 9H, tBu), 1.30-1.60 (m, 3H,  $\text{CH}_3$ ), 3.75 (s, 3H,  $\text{OCH}_3$ ), 4.35 (ddd, 1H,  $\text{H}_2$ ,  $J_{\text{H}_2\text{H}_3} = 3.5$ ,  $J_{\text{H}_2\text{NH}} = 9.2$ ,  $J_{\text{H}_2\text{F}} = 24.2$ ), 4.85 (md, 1H,  $\text{H}_3$ ,  $J_{\text{H}_3\text{F}} = 45.7$ ), 5.30 (d, 1H, NH) ; *threo* 1.45 (s, 9H, tBu), 1.20-1.60 (m, 3H,  $\text{CH}_3$ ), 3.80 (s, 3H,  $\text{OCH}_3$ ), 4.40 (dd, 1H,  $\text{H}_2$ ,  $J_{\text{H}_2\text{H}_3} = 10$ ,  $J_{\text{H}_2\text{F}} = 31.4$ ), 4.80-5.50 (m, 2H,  $\text{H}_3$  + NH).
20. 8, that was devoided of optical activity, resulted from lactamization and elimination of HF and migration of the double bond.
21. 10 was prepared from 7 by eliminating HF ( $\text{EtONa}/\text{DMF}$ ).  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ) 1.45 (s, 9H, tBu), 1.80 (d, 3H,  $\text{CH}_3$ ,  $J = 7.1$ ), 3.75 (s, 3H,  $\text{OCH}_3$ ), 5.95 (m, 1H, NH), 6.65 (q, 1H, CH=,  $J = 7.1$ ).
- We checked that 10 would have been detected in our working conditions.
22. a) H.G. VIEHE, R. MERENYI, L. STELLA, Z. JANOUZEK, Angew. Chem. Int. Ed. Engl. (1979), 18, 917 ; b) M. ANBAR, D. MEYERSTEIN, P. NETA, J. Chem. Soc. (B) (1966), 742 ; c) A.F. HEGARTY, P. O'NEILL, Tetrahedron Lett. (1987), 28, 901.
23. A. VIDAL-CROS, M. GAUDRY, A. MARQUET, submitted for publication.

(Received in France 3 November 1988)